

Phenylethanoid Glycosides from *Digitalis purpurea* and *Penstemon linarioides* with PKC α -Inhibitory Activity

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Received April 13, 1998

In a continuation of our search for potential tumor inhibitors from plants, it was found that the CH₂Cl₂–MeOH (1:1) extracts from *Digitalis purpurea* and *Penstemon linarioides* both showed PKC α -inhibitory bioactivity. Bioassay-directed fractionation of the extract from *D. purpurea* yielded the new, weakly active phenylethanoid glycoside 2-(3-hydroxy-4-methoxy-phenyl)-ethyl-*O*-(α -L-rhamnosyl)-(1 \rightarrow 3)-*O*-(α -L-rhamnosyl)-(1 \rightarrow 6)-4-*O*-*E*-feruloyl- β -D-glucopyranoside (**1**) together with the four known compounds calceolariside A (**2**), calceolariside B (**3**), forsythiaside (**4**), and plantainoside D (**5**). The extract from *P. linarioides* yielded the three known glycosides leucosceptoside A (**6**), acteoside (**7**), and poliumoside (**8**), together with the iridoid plantarenalioside (**9**). All of the isolated compounds, except compound **9**, showed inhibitory activity against PKC α with IC₅₀ values (in μ M) of 125 (**1**), 0.6 (**2**), 4.6 (**3**), 1.9 (**4**), 14.8 (**5**), 19.0 (**6**), 9.3 (**7**), and 24.4 (**8**).

As described in the previous paper in this series¹ we have added a screen for inhibitors of protein kinase C (PKC) to our yeast assays for DNA-damaging agents,² in as much as PKC has emerged as an attractive target for anticancer treatment.³ A search of plant extracts for inhibitors of PKC indicated that the detanninated CH₂Cl₂–MeOH (1:1) extracts of both *Digitalis purpurea* L. (Scrophulariaceae) and *Penstemon linarioides* Gray (Scrophulariaceae) showed PKC α -inhibitory bioactivity, and therefore bioassay-directed fractionation was undertaken on both extracts.

D. purpurea is a well-known herb with a long history of medicinal use. It is the source of the important cardiac glycosides digitoxin, gitoxin, and gitaloxin and has been used medicinally for at least 200 years.⁴ It also contains flavonoid glycosides and an anthraquinone.⁵ Recently, certain phenylethanoids such as desrhamnosyl acteoside, forsythiaside, purpureaside A, and purpureaside B have been isolated from *D. purpurea*, and it has been reported that acteoside (verbascoside) shows PKC α -inhibitory activity.⁶

P. linarioides has been reported to contain more than 50 iridoid glycosides,⁷ and various phenylethanoids such as martynoside,⁸ orbanoside,⁹ stansioside,⁹ and acteoside¹⁰ have been isolated from other plants of this genus. As noted above, acteoside has shown PKC α -inhibitory activity and also selectively inhibited aldose reductase and formation of the 5-lipoxygenase product 15-hydroxy-5,8,11,13-eicosastetraenoic acid (15-HETE) and LTB₄ in human peripheral polymorphonuclear leukocytes, as well as showing antibacterial and cytotoxic activities.¹¹

The extracts of both plants were subjected to partition between various organic solvents and aqueous MeOH, and the bioactivity of each extract against PKC α was concentrated in the *n*-BuOH fraction. For *D. purpurea* 2.52 g of crude extract gave 1.14 g of active *n*-BuOH-soluble material with an IC₅₀ of 11.7 μ g/mL against PKC α , while for *P. linarioides* 2.27 g of crude extract yielded 1.27 g of *n*-BuOH-

soluble material with an IC₅₀ of 17.1 μ g/mL against PKC α . Column chromatography of the *n*-BuOH fraction from *D. purpurea* on Si gel with the solvent CH₂Cl₂–MeOH–H₂O (8:2:0.1) gave seven fractions. The bioactive fraction 6 gave compound **1** (4.8 mg) and plantainoside (**5**) (8.0 mg) by preparative TLC on RP-18 plates with the solvent MeOH–H₂O (55:45) and repeated column chromatography on Si gel with the solvent CH₂Cl₂–MeOH–H₂O (8:2:0.1). Using the same conditions, calceolariside A (**2**, 16.8 mg), calceolariside B (**3**, 8.8 mg), and forsythiaside (**4**, 22.3 mg) were isolated from the bioactive fractions 2, 3, and 4, respectively.

In similar fashion, the *n*-BuOH fraction from *P. linarioides* was subjected to Si gel column chromatography with the solvents CH₂Cl₂–MeOH–H₂O (6:1 and 4:1). Fraction 4 yielded leucosceptoside A (**6**, 120 mg, 5.2%) by further Si gel column chromatography with the solvent CH₂Cl₂–MeOH (6:1). Fraction 6 gave acteoside (**7**, 145 mg, 6.4%) on purification by polyamide column chromatography, and fraction 8 gave poliumoside (**8**, 62.7 mg, 2.8%) and plantarenalioside (**9**, 36.8 mg, 1.6%) on Si gel column chromatography with the solvent CH₂Cl₂–MeOH–H₂O (8:2:0.1).

Compound **1** had the composition C₃₅H₅₀O₁₉ as determined by HRFABMS. Its ¹H NMR spectrum, with signals at δ 3.88 (3H, s), 7.20 (1H, d, J = 1.9 Hz), 6.81 (1H, d, J = 8.2 Hz), 7.08 (1H, dd, J = 1.9, 8.2 Hz), 6.38 (1H, d, J = 15.9 Hz) and 7.66 (1H, d, J = 15.9 Hz), and its UV spectrum, with λ_{\max} at 289 and 330 nm, suggested the presence of a feruloyl moiety, while ¹H NMR signals at δ 6.73 (1H, d, J = 2.1 Hz), 6.83 (1H, d, J = 8.1 Hz), 6.69 (1H, dd, J = 2.1, 8.1 Hz), and 2.82 (2H, br t) indicated the presence of a phenethyl moiety. Compound **1** thus belongs to the class of phenylethanoid natural products. The ¹³C NMR spectrum of **1**, in addition to signals attributable to the phenethyl and the feruloyl groups, contained signals for 18 carbons corresponding to the carbohydrate moiety. The signals in the ¹H NMR spectrum of **1** for the anomeric protons at δ 5.19 (1H, d, J = 1.4 Hz), 4.62 (1H, d, J = 1.5

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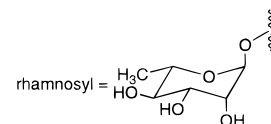
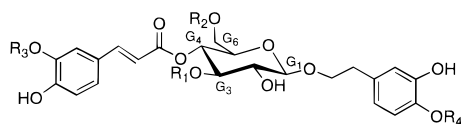
Table 1. NMR Data of Compound 1, Poliumoside (8), and Martynoside (10)

| | compound 1 | | poliumoside (8) | martynoside (10) ^a |
|----------------------------|----------------------------|------------|-----------------|-------------------------------|
| | δ_H (J value in Hz) | δ_C | δ_C | δ_C |
| aglycon | | | | |
| A-1 | | 132.8 | 131.4 | 132.7 |
| A-2 | 6.73 (d, 2.1) | 117.1 | 117.1 | 117.0 |
| A-3 | | 147.6 | 146.1 | 147.1 |
| A-4 | | 147.3 | 144.7 | 147.2 |
| A-5 | 6.83 (d, 8.1) | 112.9 | 116.3 | 112.8 |
| A-6 | 6.69 (dd, 2.1, 8.1) | 121.2 | 121.3 | 121.1 |
| A-7 | 2.82 (dt) | 72.3 | 72.3 | 72.2 |
| A-8 | | 36.6 | 36.5 | 36.5 |
| Caffeoyl- | | | | |
| C-1 | | 127.7 | 127.6 | 127.5 |
| C-2 | 7.2 (d, 1.9) | 111.8 | 115.2 | 111.9 |
| C-3 | | 149.4 | 146.8 | 150.4 |
| C-4 | | 150.8 | 149.8 | 149.1 |
| C-5 | 6.81 (d, 8.2) | 116.5 | 116.5 | 116.6 |
| C-6 | 7.08 (dd, 1.9, 8.2) | 124.4 | 123.2 | 124.2 |
| C-7 | 7.66 (d, 15.9) | 148.0 | 148.0 | 147.8 |
| C-8 | 6.38 (d, 15.9) | 115.1 | 114.7 | 115.1 |
| -COOR | | 168.0 | 168.0 | 168.3 |
| Glucosyl- | | | | |
| G-1 | 4.38 (d, 7.9) | 104.4 | 104.2 | 104.2 |
| G-2 | | 76.2 | 76.2 | 76.2 |
| G-3 | | 81.5 | 81.6 | 81.5 |
| G-4 | | 70.4 | 70.6 | 70.6 |
| G-5 | | 74.7 | 74.5 | 76.6 |
| G-6 | | 67.6 | 67.5 | 62.4 |
| Rhamnosyl-1 | | | | |
| Rh'-1 | 5.19 (d, 1.7) | 103.0 | 103.1 | 103.0 |
| Rh'-2 | | 72.3 | 72.4 | 72.3 |
| Rh'-3 | | 72.1 | 72.1 | 72.0 |
| Rh'-4 | | 73.9 | 73.9 | 73.8 |
| Rh'-5 | | 69.9 | 70.4 | 70.4 |
| Rh'-6 | | 18.4 | 18.4 | 18.4 |
| Rhamnosyl-2 | | | | |
| Rh''-1 | 4.62 (d, 1.5) | 102.3 | 102.3 | |
| Rh''-2 | | 72.3 | 72.3 | |
| Rh''-3 | | 72.0 | 72.0 | |
| Rh''-4 | | 73.7 | 73.7 | |
| Rh''-5 | | 69.9 | 69.9 | |
| Rh''-6 | | 18.0 | 18.0 | |
| CH ₃ O-(Caff-3) | 3.81 (s) | 56.5 | | 56.5 |
| CH ₃ O-(Ag-4) | 3.88 (s) | 56.4 | | 56.5 |

^a Data from Miyase et al. 12c.

Hz), and 4.38 (1H, d, 7.9 Hz) and for the terminal methyl groups at δ 1.09 (3H, d, J = 6.1 Hz) and 1.19 (3H, d, J = 6.3 Hz) indicated that **1** contained two rhamnosyl units and one glucosyl unit.

Comparison of the NMR data of **1** with those of the known compound poliumoside (**8**) (Table 1) indicated that both carbon and proton chemical shifts of the sugar part of **1** were very similar to those of poliumoside (**8**). Compound **1** also had very similar ¹³C NMR data to those of martynoside **10**,¹² after making adjustments for the fact that **10** has one rhamnose less than compound **1**. A careful comparison of the ¹³C NMR data of **1** with the data of **10** showed that the only significant differences occurred at G₅ and G₆ in **1**, thus suggesting that the additional rhamnosylation in **1** took place at the G₆ position of **10**.¹² The long-range couplings between H-G₄ (δ_H 4.99) and -COOR (δ_C 168.0) in the HMBC spectrum of **1** indicated that the feruloyl group was connected at G₄ via an ester linkage, and the long-range couplings between H-Rh₁' (δ_H 5.19) and C-G₃ (δ_C 81.5) and between H-Rh₁' (δ_H 4.62) and C-G₆ (δ_C 67.6) demonstrated that one rhamnosyl group was located at the G₆ and the other at the G₃ position. The correlation between H-G₁ (δ_H 4.38) and C- α (δ_C 72.3) indicated the location of the α -phenylethanyl glucoside moiety.



| | | | | |
|------------------------------|-----------------------------|-----------------------------|------------------------------------|----------------------------------|
| 1 | R ₁ = rhamnosyl, | R ₂ = rhamnosyl, | R ₃ = CH ₃ , | R ₄ = CH ₃ |
| 2 (Calceolarioside A) | R ₁ = H, | R ₂ = H, | R ₃ = H, | R ₄ = H |
| 4 (Forsythiaside) | R ₁ = H, | R ₂ = rhamnosyl, | R ₃ = H, | R ₄ = H |
| 6 (Leucosceptoside) | R ₁ = rhamnosyl, | R ₂ = H, | R ₃ = CH ₃ , | R ₄ = H |
| 7 (Acteoside) | R ₁ = rhamnosyl, | R ₂ = H, | R ₃ = H, | R ₄ = H |
| 8 (Poliumoside) | R ₁ = rhamnosyl, | R ₂ = rhamnosyl, | R ₃ = H, | R ₄ = H |
| 10 (Martynoside) | R ₁ = rhamnosyl, | R ₂ = H, | R ₃ = CH ₃ , | R ₄ = CH ₃ |

In a NOESY spectrum, NOE correlations observed between the CH₃O- signal at δ_H 3.81 and the proton signal at δ_H 7.20 (1H, d, 2.1) showed this CH₃O- group to be located at the C-3 position of the feruloyl moiety. Similarly, the other CH₃O- group could be assigned to the C-4 position of the phenethyl group by the correlation between the CH₃O- signal at δ_H 3.88 (3H, s) and proton signal at δ_H 6.83 (1H, d, J = 8.1 Hz). Based on these data, **1** could be assigned as 2-(3-hydroxy-4-methoxyphenyl)-ethyl-*O*-(α -L-rhamnosyl)-(1 \rightarrow 3)-*O*-(α -L-rhamnosyl)-(1 \rightarrow 6)-4-*O*-*E*-feruloyl- β -D-glucopyranoside.

Compounds **2**–**9** were assigned as calceolarioside A,¹³ calceolarioside B,¹³ forsythiaside,¹⁴ plantainoside D,¹⁵ leucosceptoside,¹² acteoside,^{9,12,13,16} poliumoside,¹⁷ and plantarenalioside,¹⁷ respectively, by comparison of their ¹H and ¹³C NMR, DQCOSY, HMQC, HMBC, and NOESY spectra with literature data. They also showed the same HRFABMS, [α]_D, UV, and IR data with values in the literature. Calceolarioside A and plantarenalioside were also identified by direct comparison with authentic samples. Plantainoside D was isolated for the first time from *P. linarioides*.

All of the isolated compounds except compound **9** showed inhibitory activity against PKC α , with IC₅₀ values (μ M) of 125 (**1**), 0.6 (**2**), 4.6 (**3**), 1.9 (**4**), 14.8 (**5**), 19.0 (**6**), 9.3 (**7**) and 24.4 (**8**). The PKC α inhibitory bioactivities of **1**–**6** and **8** have not been reported previously in the literature.

Experimental Section

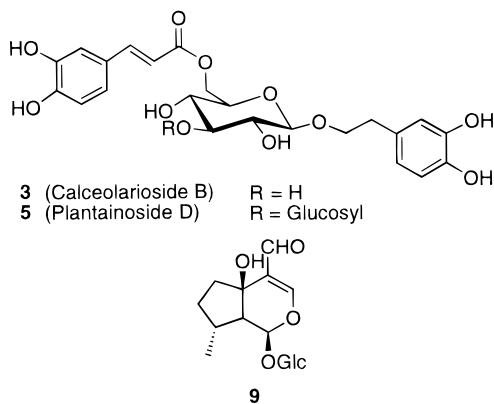
General Experimental Procedures. Optical rotations were recorded with a Perkin-Elmer 241 Polarimeter. UV spectra were measured on a Beckman DU-50 instrument and IR spectra on a Nicolet Impact 400 spectrophotometer. NMR spectra were recorded in CD₃OD on a Varian Unity 400 NMR instrument at 399.951 MHz for ¹H and 100.578 MHz for ¹³C, using standard Varian pulse sequences. Exact mass measurements were obtained at the Nebraska Center for Mass Spectrometry. Other conditions were as previously described.²

Plant Material. Stems, leaves, and fruit of *Digitalis purpurea* L. (Scrophulariaceae) were collected on Norfolk Island in May 1965 (PR-9503, B633363), and the whole plant of *Penstemon linarioides* Gray (Scrophulariaceae) was collected in New Mexico in June 1964 (PR-8513, B632569). Voucher specimens are on deposit in the Herbarium of the National Arboretum, Agricultural Research Service, U. S. D. A., Washington, DC. The dried samples were extracted with CH₂Cl₂-MeOH (1:1) to give 2.52 g of *D. purpurea* extract as HEX 652 and 2.27 g of *P. linarioides* extract as HEX 1216.

Isolation of Phenylethanoids 1–5. The CH₂Cl₂-MeOH extract HEX 652 (2.52 g) from *D. purpurea* was passed through a polyamide column (80 g, ICN Pharmaceuticals, Inc., Eschwege, Germany) to yield a detannated fraction (2.16 g, PKC α IC₅₀ = 78 μ g/mL). After partition of the detannated fraction between EtOAc and aqueous MeOH, followed by partition of the aqueous MeOH fraction between H₂O and

n-BuOH, a bioactive *n*-BuOH fraction (1.14 g, PKC α IC₅₀ = 12 μ g/mL) was obtained. This was subjected to column chromatography on Si gel (60 g) with elution by CH₂Cl₂-MeOH-H₂O (8:2:0.1) to give seven fractions after combination of similar components as determined by TLC. Purification of active compounds from the bioactive fractions was achieved by preparative TLC on RP-18 with MeOH-H₂O (55:45) followed by column chromatography on Si gel with CH₂Cl₂-MeOH-H₂O (8:2:0.1). Fraction 2 yielded calceolarioside A (**2**, 16.8 mg, 0.67%); fraction 3 gave calceolarioside B (**3**, 8.8 mg, 0.35%); fraction 4 gave forsythiaside (**4**, 22.3 mg, 0.89%); fraction 6 gave plantainoside D (**5**, 8.0 mg, 0.32%) and the new natural product **1** (4.79 mg, 0.19%). The structures of compounds **2-5** were assigned by ¹H and ¹³C NMR, DQCOSEY, HMQC, HMBC, and NOESY spectra, and by HRFABMS, [α]_D, UV, and IR. Calceolarioside A was also identified by direct comparison with an authentic sample.¹³

Isolation of Phenylethanoids 6-9. The CH₂Cl₂-MeOH (1:1) extract from *P. linarioides* was partitioned between EtOAc and aqueous MeOH, and the active aqueous MeOH fraction was then partitioned between H₂O and *n*-BuOH. The bioactive *n*-BuOH fraction (1.27 g, 56.1%, with IC₅₀ = 17.1 μ g/mL against PKC α) was subjected to column chromatography on Si gel with the solvent CH₂Cl₂-MeOH (6:1 and 4:1) to give 10 fractions, of which fractions 4, 6, and 8 showed PKC α -inhibitory activity. Leucosceptoside A (**6**, 120 mg, 5.2%) was isolated from fraction 4 by further Si gel column chromatography with the solvent CH₂Cl₂-MeOH (6:1). Acteoside (**7**, 145 mg, 6.4%) was purified by polyamide column chromatography of fraction 6, and pliumoside (**8**, 62.7 mg, 2.8%) and plantarenalioside (**9**, 36.8 mg, 1.6%) were obtained by Si gel column chromatography of fraction 8 with the solvent CH₂Cl₂-MeOH-H₂O (8:2:0.1). The structures of compounds **6-8** were assigned by comparison of ¹H and ¹³C NMR, NOESY, HMQC and HMBC spectra, and UV, MS, and [α]_D values with literature data; all data were identical to those published.^{10,12,14,15,19} Plantarenalioside (**9**) was identified by direct comparison with an authentic sample (co-TLC, ¹H and ¹³C NMR).



Compound 1: light yellow gum-like substance, [α]_D²³ -60.4° (c 0.22, MeOH), UV (MeOH) λ _{max} (log ϵ) 218 (4.06), 235 sh (3.98), 289 (3.85), and 330 (4.05) nm; IR ν _{max} (KBr) 3600-3100 (OH), 1720 (conjugated COOR), 1600 (>C=C<); ¹H and ¹³C NMR data see Table 1; FABMS *m/z* 805.3077 (calcd for C₃₇H₅₀O₁₉Li, 805.3106).

PKC α -Inhibitory Bioassay. Bioassay for inhibition of PKC was carried out as described previously.¹

Acknowledgment. This work was supported by a National Cooperative Drug Discovery Group award to the University of Virginia (U19 CA 50771, Dr. S. M. Hecht, Principal Investigator), and this support is gratefully acknowledged. The authors thank Dr. Corrado Galeffi, Laboratorio di Chimica del Farmaco, Rome, Italy, for an authentic sample of calceolarioside A. Mass spectra were obtained by Mr. Kim Harich, Virginia Polytechnic Institute and State University, and the Nebraska Center for Mass Spectrometry.

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NP980147S